

(An ISO 3297: 2007 Certified Organization) Vol. 3, Issue 2, February 2014

Ultra Structural Changes in the Male Reproductive Organ of *Eisenia foetida* after Treatment with an Organophosphorus Pesticide

K. Bansiwal¹, N. Rai²

Research Fellow, Department of Environmental Sciences, M. L. Sukhadia University, Udaipur, Rajasthan, India¹

Associate Professor, Department of Environmental Sciences, M. L. Sukhadia University, Udaipur, Rajasthan, India²

Abstract: Malathion is an organophosphate insecticide and used as a test chemical to assess its toxicity for male reproductive organ of *Eisenia foetida*. Worms exposed to high dose (12000mg/Kg soil of 5 percent Malathion) for 96 hours showed significant changes in ultra structure of male organ of *E. foetida*. These changes include vacouliged cytophore and spermatids, deshaped mid piece of spermatid. It is suggested that Malathion at 12000mg/Kg soil for 96 hours has caused deformities at ultra structure level in male reproductive organ of *E. foetida* that will ultimately lower the reproductive rate of *E. Foetida*. Thus it can be said that Malathion is not only a neurotoxin agent but also a cytotoxic agent that also affects the other organs apart from nervous system.

Key words: Malathion, testes spermatid, ultra structure

I. INTRODUCTION

Organophosphates are insecticides, exerting their toxicity by binding to acetyl cholinesterase, inhibiting the action of this enzyme (which degrade the neurotransmitter acetylcholine), and eventually killing the organisms ([1], [2], [3]). Organophosphates are frequently used as pesticides due to their low half life. Malathion has been used for present investigation since it is commonly used in agriculture for killing sucking and chewing insect etc.

Malathion is considered one of most innocuous organophosphates for humans, which led to an indiscriminate use of this pesticide everywhere.

Among soil organisms, earthworms are particularly selected for the assessments because of these organisms play a vital role in vermi-composting and nutrient cycling and they also constitute a big part of the invertebrate biomass in the soil. Earthworms are the most suitable organism for use as key bioindicators organisms for testing the toxicity of chemicals in soil ([4], [5], and [6]) and also adopted as a standard organism for ecotoxicological testing ([7], [8], [9]). Furthermore, among the various earthworm species, *Eisenia foetida* is especially appropriate for the toxicity tests because it can be easily reared on a variety of organic wastes with short lifecycle. Its susceptibility to chemicals resembles that of true soil organisms. Sensitivity tests reveal that *E. foetida* is comparatively less sensitive than other earthworm species ([10], [11], and [12]). It is a suitable test model species for toxicological experiments [13].

Malathion-induced morphologic alterations have been found in the mouse testes [14]. According to reference [15] in laboratory rats the organophosphate causes a decrease of testicular mass with degenerative injuries and partial failing of the spermatogenesis and an increase in abnormalities of spermatozoa. In birds, Malathion induces germ cell degeneration possibly by altering cholinergic functions [16]. Worms makes direct contact with the ground and absorbs pesticides both from skin and digestive system, thus the pollutants are absorbed 5 to 10 times.

The toxic effects on earthworm gametes should correspond directly to their population and indirectly to other wildlife [17]. The present work evaluates the changes produced after treatment of Malathion in the male reproductive organ of *Eisenia foetida* at intra cellular level discussed in the form of electron micrographs.



ISSN: 2319-8753

International Journal of Innovative Research in Science, Engineering and Technology

(An ISO 3297: 2007 Certified Organization)

Vol. 3, Issue 2, February 2014

II. MATERIALS & METHODS

- 1. Experimental model
- 2. Pesticide
- 3. Experimental set-up
- 4. Treatment of worms
- 5. Electron microscopy of clitellum
- 1. Experimental Model: Earthworms, *E. foetida* was chosen to evaluate toxicity in present study because it was easy to handle and its easy availability. Earthworms were procured from the vermicomposting unit of Rajasthan College of Agriculture, Udaipur. They were maintained in the laboratory condition, after 15 days acclimatization they were used for further conducted experiment. The worms used in the experiment were of approximately same body weight and body length.
- 2. Chemical: The pesticide used in the experiment was Malathion (5%). It was purchased from the local market's shop. The LD50 value of this pesticide for the *E. foetida* was evaluated by author first that is 16000mg/kg of soil and the result of this LD50 supported by the reference [18].
- **3.** Experimental set-up: Our method of the experiment was based on the method as in reference [19]. We used plastic tub for our experiment. Dried soil (from nearby farmland) was crusted and filtered through a fine mesh sieve. One kg of fine soil was than poured in each plastic tub and water was added to moistened the soil and 250 mg dried powdered (3 week old) cow dung was also added to each plastic tubs. Cow dung was added to avoid starvation. 20 mature earthworms (some age group) were added to each plastic tub. Each plastic tub was covered with muslin cloth. Thus one control set and one experimental set were prepared 3 replicates were used for each set.
- 4. Treatment of the worm with Malathion: Higher sub lethal lose of Malathion (12000mg/kg soil) was selected to study the toxicity of Malathion for *E. foetida*. Thus in experimental set higher sub lethal dose of Malathion was added and mixed throughoughtly in the plastic tub. Regular water supply was done to maintained 60% to 70% moisture level. After 96 hours of the treatment the worms were taken from the plastic tub and dissected out for desired organ of the body.
- 5. Electron microscopy of Testes: Worms were dissected from 9 to 11 segment from both the group (control and experimental) and immersed in Karnovsky fixer (1% glutaraldehyde and 4% paraformaldehye in 0.2 M (cacodylate buffer at pH 7.4). Then washed in 0.2M Cacodylate buffer (pH 7.4) and post fixed in 1% Osmium tetroxide, similarly buffered, for 1h following dehydration in an acetone series, the tissue were embedded in resin. Ultra thin sections were stained with uranyl acetate followed by lead citrate, examined and photographed with a Morgagni 268D, Transmission Electron Microscope.

III. RESULTS AND DISCUSSION

In *Eisenia foetida*, spermatogenesis begins in testes where spermatogonia divide mitotically and with an incomplete cytokinesis originating a common cytoplasmatic mass. The cytophore which connects polarizes and synchronizes the surrounding cells through cytoplasmic bridges. From there, the whole spermatogonia move to the seminal vesicles where spermatogenesis process is completed.

In the lumen of the vesiculae seminalis, cytophore, coelomcytes, and different stages of developing spermatocytes, and different stages of developing spermatozoids are floating. A central common nutrient anucleate mass of cytoplasm is known as cytophore. It consists of numerous mitochondria, clusters of ribosomes, and elaborate stack of parallel smooth endoplasmic reticulum, rough endoplasmic reticulum cisternae, membrane-bound dense bodies and degenerated dictyosomes (Fig.1 & 2).



(An ISO 3297: 2007 Certified Organization)

Vol. 3, Issue 2, February 2014



Fig.1: Transmission electron micrograph of the cytophore of a control worm. It contains parallel cisternae of smooth endoplasmic reticulum (ER), rough endoplasmic reticulum (rER), ribosomes (R) and mitochondria (M). 8900x



Fig.2: Transmission electron micrographs of the cytophore of a control worm. It contains parallel cistenae of smooth endoplasmic reticulum (ER), rough endoplasmic reticulum (rER), ribosomes (R) and mitochondria (M). 5600x

All spermatic cells and the developing spermatids are around the cytophore through a cytoplasmic connection called zonula collaris (Fig.3).



Fig.3: Immature spermatids are attached to the cytophore (C) of a control worm through the zonula collaris (ZC). Each spermatid (S) has a large nucleus (N). 1100x

After maturation spermatids get detached from the cytophore and move freely in the coelom, thereafter the cytophore becomes degenerated (Fig.4 and 5) Electron microscopic observation of vesiculae seminalis of worms treated with 12 gm/kg Malathion shows a large no. of abnormalities confined to cytophore and different stages of spermatic development.



(An ISO 3297: 2007 Certified Organization)

Vol. 3, Issue 2, February 2014



Fig.4: TEM of a degenerated cytophore (C) a control worm, after the detachment of the mature spermatids (mS). 2200x



Fig.5: TEM of mature spermatids (S) of a control worm after complete detachment from related cytophore. (N=large nucleus). 3500x

Electron-micrograph number 6, 7 (Fig.6 & 7) clearly shows a highly vacuolised cytophore of the worms exposed to a higher sub lethal dose of Malathion (12gm/kg). Abnormal cytophore contained a large number of vacuoles of various sizes (Fig.6, 7). One of the adverse effect of Malathion on spermatids have also observed that was presence of large vacuoles within the mature spermatid (Fig. 8) although whereas mature spermatid of the worm from control group, did not reveal any such abnormality (Fig.11). Deshaped, elongated spermatid was also seen in the treated worm (Fig.9 & 10).



Fig.6: TEM of abnormal cytophore (C) after 12gm/kg Malathion treatment, showing high number of vacuoles (V). 3500x



(An ISO 3297: 2007 Certified Organization)

Vol. 3, Issue 2, February 2014



Fig.7: TEM of abnormal cytophore after 12gm/kg Malathion treatment, showing high number of vacuoles (V). 3500x



Fig.8: Transverse sections of spermatids after 12 mg/kg Malathion treatment, showing vacuoles (v) within spermatid. 2200x



Fig.9: Longitudinal sections of spermatids showing vacuoles and deshaped structure, (showing by arrows) after treatment with 12gm/kg Malathion. 2800x



Fig.10: Longitudinal sections of spermatids showing vacuole (V) and deshaped mid piece (dM) (showing by arrows). 3500x



(An ISO 3297: 2007 Certified Organization)

Vol. 3, Issue 2, February 2014



Fig.11: Longitudinal sections of spermatids of control worm, Eisenia foetida.1000x (MI = middle piece, N = nucleus)

Invertebrates are frequently used as biomarkers for the possible effects of xenobiotics applied to the environment [20]. Earthworms have been successfully used as sentinels of soil and also for non target toxicity tests of chemicals used in agricultural practices [21].

Currently, there is much concern about the adverse effects of environmental chemical agents in biological systems [22]. Malathion is lethal to many living systems due to its indiscriminate utilization in different fields like, agriculture, houses and gardens. Malathion affects the neuromuscular function, altering the balance of Ca^{++} and K^+ which seems to be the factor responsible for changes in the morphology and activity of the earthworms. This would explain the phenomenon of coiling of the tail, observed in all individual treated with Malathion [18].

Previously many authors ([23], [24], and [25]) have performed a detailed study on spermatogenesis of *Eisenia foetida* including light and electron microscopy. These studies include information on the cytophore and gonial development. And later have been summarized by reference [26]. The present study gives concrete pictorial evidence that Malathion causes abnormality in the ultra structure of cytophore as well as affects spermatids of *Eisenia foetida*. These evidences are well supported by the references [27], [28], [29].

Other studies report that spermatozoa could be damaged by the effect of pesticide on spermatogonical morula, altering the ultra structure of the cytoskeleton of the cytophore (common cytoplasmic mass), affecting therefore the normal development of the spermatogenesis and the spermiogenesis [17]. Malathion has an acute and direct cytotoxic effect (cellular death) on human lymphocytes, without causing damage to the DNA, but their active metabolites, malaoxon and isomalathion, could act later by breaking DNA chains and Malathion degraded to its active metabolites malaoxon and isomalathion, which could also alter spermatogenesis ([18],[30]). According to references [31], [32], [33] and [18], *E. foetida* treated with Malathion exhibited a completely disordered structure with loss of central hillus, vacuolization and cells with small and pyknotics nuclei.

This emphasizes the idea that Malathion alters the testicular function seriously in this species, as reported earlier for earthworms and mice treated with other organophosphates pesticides.

Malathion has a direct cytotoxic effect causing coiling of the tail, with increase of metachromasia of the chromatin of the spermatozoa and altering the sperm count and also affects testicular structure, with increased vacuolization and hyperpyknosis of the nuclear of spermatogonia [18].

Damage to spermatozoa can be of particular importance since it may cause adverse effects that may not be "visible" in terms of the morphology of the organism or male fertility but in terms of fetal loss and congenital malformations ([34], [35]).

According to ref. [36], in mice it has been reported that Malathion was a potent cell cycle inhibitor that induces DNA damage and is capable of interfering with DNA replication; this suggests that Malathion is a genotoxic agent and may be regarded or a potential germ cell mutagen.

The main objective in the ecotoxicological work is to provide information to estimate ecological risk [37], but the relative inability to identify critical endpoints remains a fundamental problem. According to ref. [38], features to study in this regard are the biochemical, cellular and physiological effects. Our study evidently confirms that the information provided at the ultra cellular level by ultra structural changes during spermatogenesis in *E. foetida* could well in future fulfils the above criterion.

It can be concluded from this electron microscopic investigation of male reproductive organ of *E. foetida* after exposure of Malathion that, it is a cytotoxic agent that alters reproductive capability of *E. foetida* and ultimately reduce the reproductive out put of this worm.



ISSN: 2319-8753

International Journal of Innovative Research in Science, Engineering and Technology

(An ISO 3297: 2007 Certified Organization)

Vol. 3, Issue 2, February 2014

REFERENCES

- [1] T.R. Fokuto, "Mechanism of action of organophosphorus and carbamate pesticides," Environ. Health Perspect., vol. 87, pp. 245-254, 1990.
- [2] M. O'Malley, "Clinical evaluation of pesticide exposure and positioning," *Lancet.*, vol. 349, pp. 1161-1166, 1997.
- [3] Y. Solberg, and M. Belkin, "The role of excitotoxicity in organophosphorous nerve agents central poisoning," *Trends Pharmacol. Sci.*, vol. 18, pp. 183-185, 1997.
- [4] C.A. Challahan, "Earthworm as ecotoxicological assessment tools. *In*: (Eds.: Edward, C.A. and Neuhauser, E.F.). Earthworms in waste and environmental assessment. SPB Academic, The Hague. pp. 295-301, 1988.
- [5] Goats, G.C. and Edwards, C.A.: Earthworms in waste and environmental assessment. *In:* The prediction of field toxicity of chemicals to earthworms by laboratory methods. (Eds.: Edwards, C.A., Neuhauser), E.F. SPB Academic, The Hague, pp.283-294, 1988.
- [6] A. Bouche, "Ecotoxicology of earthworms. In: Greig-smith Earthworm species and ecotoxicological studies (Eds.: P.W., Becker, H. Edwards, P.J. and Heimbach, F.). Intercept, Andover. pp. 20-35, 1992.
- [7] EEC (European Economic Community). Directive 79/831/EEC, Annex V, Part C. Method for the determination of ecotoxicity level 1. Earthworms: artificial soil test. Commission of the European Communities, DGX1/128/82, Rev. 5, Brussels. (1984).
- [8] OECD.: Earthworm, acute toxicity tests. In: Organization for economic Co-operation and Development (Eds.) OECD guidelines for testing of chemicals, Paris, (1984).
- [9] OECD, Guidelines for testing of chemicals: earthworm reproduction test (Eisenia foetida) OECD, Paris, 2000.
- [10] W.C. Ma, and J. Bodt, "Differences in toxicity of the insecticide chloropyrifos to six species of earthworms (Oligochaeta, Lumbricidae) in standardized soil tests," *Bull. Environ. Contain. Toxicol.*, vol. 50, pp. 864-870, 1993.
- [11] D.G. Fitzgerald, K.A. Warner, R.A. Lanno, and D.G. Dixon, "Assessing the effects of modifying factors on pentachlorophenol toxicity to earthworms: application of body residues," *Environ. Toxicol. Chem.*, vol. 15, pp. 2299-2304, 1996.
- [12] H. Kula, and O. Larink, "Development and standardization of test methods for the pesticide side effects on earthworms," Acta. Zool. Fenn., vol. 196, pp. 338-341, 1995.
- [13] D. S. Spurgeon, and S. P. Hapkin, "Effect of metal contaminated soil on the growth, sexual development and early cocoon production of the earthworm, Eisenia foetida, with reference to zinc," *Ecotoxicol. Environ. Saf.*, vol. 35, pp. 86-95, 1996.
- [14] H. Contreras, and E. Bustos-Obregon, "Morphological alteration in mouse testes by a single dose of Malathion," J. Exp. Zool., vol. 284, pp. 355-359, 1999.
- [15] M. Atef, S. Youssef, A. Ramadan, M. Nawito, E. I. M. Sayed, and H. Rehman, "Influence of Phoxim on testicular and seminal vesical organs, testosterone and cholinesterase level and its tissue residues in male rats," *Deutrch Tierarztl. Wochenschr.*, vol. 8, pp. 301-305, 1995.
- [16] S. Maitra, and R. Sarkar, "Influence of methyl parathion on gametogenic and acetylcholinesterase activity in the testers of white throated munia (Lonchura Malabarica)," Arch. Environ. Contam. Toxicol., vol. 30, pp. 384-389, 1996.
- [17] J. Sorour, and I. Larink, "Toxic effects of benomyl on the ultrastructure during spermatogenesis of earthworm *Eisenia foetida*," *Ecotoxicol. Envion Saf.*, vol. 50, pp. 180-188, 2001.
- [18] E. N. Omar, and E. O. Bustos, "Sublethal doses of Malathion alter male reproductive parameter of Eisenia foetida," J.Morph., vol. 22(4), pp. 297-302, 2004.
- [19] S. Yasmin, and D. Souza, "Effect of pesticides on reproductive output of *Eisenia foetid*,". *Bull.Env. Contam. Toxicol.*, vol. 79, pp. 592-532, 2007.
- [20] J.M. Weeks, C. Spurgon, P.K. Hankard, J.E. Kammenga, R. Dallinger, H. Kohler, V. Simonsen, and N. Scott-fordrmand, "Critical analysis of soil invertebrate biomarkers: a field case study in Avonmouth," UK. *Ecotoxicol.* vol. 13, pp 817-822, 2004.
- [21] L.H. Booth, S. Hodge, and K. O'Halloran, "Use of biomarkers in earthworms to detect use of field application of a model organophosphate pesticide," Arch. Environ. Cont. Toxicol., vol. 67, pp. 633-640, 2001.
- [22] J. Toppari, J.C. Larson, P. Christiansen, and A. Giwercman, "Male reproductive health and environmental xenoestrogens," *Environ. Health Perpect.*, vol. 104, pp. 741-803, 1996.
- [23] C. Stang-Voss, and J. Stavbesand, "Uber die Neubildung Von Mitochondrien: Elektronen mikroskopische Untersuchungen an spermatiden Von Eisenia foetida (Annelidae," Z. Zellforsch., vol. 111, pp. 127-142, 1970.
- [24] C. Stang-Voss, "Zur Entstehung des Golgi-Apparater: Eloktronen-Mikroskopische Untersuchungen an Spermatiden Von Eisenia foetida (Annelida)," Z Zellforsch., vol. 109, pp. 287-296, 1970.
- [25] B.G.M. Jamieson, The Ultrastructure of the Oligochacta. Academic press. London, 1981.
- [26] Jamieson, B.G.M.: Oligocheata. In: 'Microscopic Anatomy of Invertibrates''. Vol.7, "Annelida" (Eds.: F.W. Harrison and S.L.Gardiner). Wiley, New York, pp.217-322, 1992.
- [27] B. Senapati, J. Beiswa, S. Pani, and S. Sahu, "Ecotoxicological effects of Malathion on Earthworms," Soil Biol. Biochem., vol. 24, pp. 1719-1722, 1992.
- [28] C. Bauer, and A. Rombke, "Factors influencing the toxicity of two pesticides on three Lumbricid species in laboratory test" Soil Biol. Biochem., vol. 29, pp. 705-708, 1997.
- [29] E. N. Omar, and E. O. Bustos, "Effect of Malathion on the male reproductive organ of Eisenia foetida," Asian. J. Androl., vol. 7(1), pp. 97-101, 2005.
- [30] J. Blasiak, P. Jaloszynski, A. Trzeciaks, and K. Szyfter, "In vitro studies on the genotoxicity of the organophosphorus insecticide Malathion and its two analogues," *Mutat. Res.*, vol. 445, pp. 275-283, 1999.
- [31] C. Sobargo, and E. Bustos-Obregon, "Acute effect of parathion on the seminiferous epithelium of immature mice," *Rev. Chil. Anat.*, vol. 18, pp. 61-68, 2000.
- [32] E. Bustos-Obregon, and R. Goicochea, "Pesticide soil contamination mainly affects earthworm male reproductive parameters," Asian J. Androl., vol. 4, pp. 195-199, 2002.



(An ISO 3297: 2007 Certified Organization)

Vol. 3, Issue 2, February 2014

- [33] Bustos-Obregon, and P. Gonzalez Harmazabal, "Effect of a single dose of malathion on spermatogenesis in mice," Asian J. Androl., vol. 5, pp. 105-107, 2003.
- [34] A. Sophie, and A. J. Reinecke, "The influence of lend and manganese on spermatozoa of *Eisenia foetida* (Oligochaeta)," *Soil-Biol. Biochem.*, vol. 29, pp. 737-742, 1997.
- [35] T.S. Thompson, R.G. Treble, A. Magliocco, J.R. Roettger, and J.C. Eichhorst, "Case study: fetal poisoning by Malathion," Forensic Sci. Int., vol 95, pp. 89-98, 1998.
- [36] S. Giri, S.B. Prasad, A. Giri, and D. Sharma, "Genotoxic effects of malathion: an organophosphorous insecticide, using three mammalian bioassay in viva," *Mutat, Res.*, vol. 514, pp. 223-231, 2002.
- [37] R. Wilson, and E.A.C. Crouch, "Risk assessment and comparisons: an introduction science," vol. 236, pp. 267-270 1987.
- [38] M. Gilbertson, "Fresh water avian and mammalian predators as indicators of aquatic environment quality," *Environ Monitoring Assessment.*, vol. 15, pp. 219-223, 1990.