

(An ISO 3297: 2007 Certified Organization) Vol. 3, Issue 6, June 2014

Early Evaluation of Etilological Treatmetn in Young Adults, Chronically Infected *Trypanosama Cruzi* Detected Using Polymerase Chain Reaction

K.Sathish¹, Dr.S.Niraimathi¹, S.Justin Jayaraj², K.Kamalakannan³

¹PG Research, Department of Biochemistry, RVS College of Arts & Science, Karaikal, India.

²Department of Pharmaceutical Analysis, E.G.S.Pillay College of Pharmacy, Nagapattinam, India.

³Department of Science & Humanities, Arasu Engineering College, Kumbakonam, India.

ABSTRACT: Fourteen patients (age limits: 18-30) chronically infected with Trypanosoma cruzi and presenting positive blood PCR reactions, were submitted to etiological treatment with benznidazole, in Nagapattinam district rural an area. A control group of five patients of the same age and positive by PCR did not receive treatment. Post-treatment follow-up was performed with PCR and conventional serology. PCR became negative in 12/14 of the treated patients after six months follow up, compared to 1/5 in the control group (p = 0.01). All patients remained serologically positive after treatment. The reduction of PCR signals of infection after treatment may become an early evidence of cure in chronically infected young adult patients.

I. INTRODUCTION

Chagas disease, caused by the parasite *Trypanosama cruzi*, is transmitted to humans and animal by insects (vectors) located in America, mainly in rural areas of Latin America. Humans can be infected by blood transfusions or tissue and organ transplants from infected donors, from a mother to her child (congenital transmission), orally by consuming contaminated food with faeces from infected insects, or by accidental exposure in the laboratory. Additionally, this disease has extended, due to migrations, to countries such as Spain (6) and the USA (7). The etiological treatment of chronically infected patients has been recommended by the World Health Organization (10), Brazil (8) and Argentina (9) treatment guidelines also recommend treatment for such patients, stating that the decision should be taken by each patient and his/her physician. Conventional serologic (Enzyme Linked Immunosorbent Assay - ELISA, Indirect Hemagglutination - IHA) and parasitologic (Hemoculture, Xenodiagnosis) methods are recommended for the control of therapeutic efficacy. However, in the chronic phase, conventional reactions only become negative years after treatment (1, 10), and conventional parasitologic methods have low sensitivity (4). The Polymerase Chain Reaction (PCR), detecting Trypanosoma cruzi DNA (13), showed higher diagnostic sensitivity than conventional methods in chronic Chagas disease patients (14). In this work, we explored the usefulness of PCR as a tool for early evaluation of the efficacy of etiologic treatment in young adults infected by T. cruzi. Treatment was also controlled with conventional serology (IHA, ELISA)

Subjects

II. MATERIALS AND METHODS

Ninety-one patients were interviewed, age ranged from 18 to 30 years old, and they had two positive serological tests (ELISA and IHA). The PCR test was applied to all of them. The reaction was positive in 36/91 (65.5%). Treatment was initiated in 18 patients and 14 completed the whole treatment course. The control group was composed of five patients who rejected treatment or received it after this study. Out of 19 patients thirteen were women. All lived inNagapattinam rural, an area with no Victorial transmission, and signed an informed consent. Exclusion criteria were: previous treatment for T.

Copyright to IJIRSET



(An ISO 3297: 2007 Certified Organization)

Vol. 3, Issue 6, June 2014

cruzi, pregnancy, liver or kidney disease, low leukocyte or platelet counts, alcoholism, severe pathology associated to Chagas disease and breast-feeding. Treatment Two groups were studied the treated group (TG), included 14 patients (age range 23.6 ± 3.5 years), which received benznidazole at 5 mg/kg/day for 60 days, by oral route. The control group (CG), included 5 patients (age range 26.6 ± 4.9 years), which did not receive treatment. The follow-up was of 6 months post-treatment for TG and from 2 weeks to 6 months for CG. Serology After separating the serum, IHA and ELISA tests were done, using Wiener Laboratory kits (government research area) and following the recommended procedures and cut-off points (1/16 dilution for IHA and 0.22 absorbance for ELISA)

Polymerase Chain Reaction (PCR)

The procedure used by Britto et al. (3) with minor modifications was used. Blood (5 mL) was diluted 1:1 in 6 M-Guanidine EDTA 0.2 M buffer. The sample was heated at 100° C 10 minutes. DDNA was extracted in minipreps using 100 µsample aliquots, using phenol-chlorophorm-isoamilic alcohol and sodium acetateethanol precipitation. Amplification mixtures included primers 121 (5'-AAA TAA TGT ACG GGT GAG ATG CAT GA-3') and 122 (5'-GGT TCG ATT GGG GTTGGT GTA ATA TA-3'). Amplification was carried out in a thermal cycler (M. J. Research, Watertown, Masssachusetts, USA) with the following cycle sequence: cycle 1(x2) 1 min at 98° C and 2 min at 64° C; cycle 2 (x33): 1 min at 94 °C and 2 min at 64° C; cycle 3 (x1): 1 min at 72° C and 2 min at 25° C. Amplified productswere electrophoresed in 0.2 % agarose minigels, stained with ethidium bromide and revealed with ultraviolet light. Positive samples displayed a 330 bp band. Because cross-contamination or PCR artifacts are a constant risk, a maximum of 5 samples was processed, together with a negative and a positive control, per extraction session. PCR mixtures and DNA extraction were performed in separate chambers. All reagents were prepared in aliquots, using exclusive pipettes and filter tips. Amplicons were electrophoresed in a separate room.

Statistical analysis

The significance of differences between proportions was calculated with the Fisher's exact test.

III. RESULTS

The main outcomes of ELISA, IHA, and PCR for each patient are shown in Tables 1 and 2. PCR became negative in 12/14 (85.7%) TG patients at month 6 post-treatment, and in 1/5 (20%) CG patients (p = 0.01). All patients remained serologically positive after treatment (TG), or in the follow-up of the CG. Seven of 14 (50%) patients in the TG presented adverse effects of benznidazole. These were: allergic dermatitis (4/7; 57.2%), peripheral polyneuritis (2/7; 28.6%), and asthenia (1/7; 14.3%). Nevertheless, treatment could be completed in all patients, by either diminishing temporarily the dose or by symptomatic medication. In two cases, adverse effects disappeared spontaneously after the first days of treatment.

Table 1. Result of serological tests and PCR before/ after treatment with benznidazole of young adults, infected with Trypanosoma cruzi .

Patie	Before treatment			After treatment		
nts						
	IHA	ELIS	PC	IHA	ELI	PC
		А	R		SA	R
1	1/1024	+ve	+ve	1/1024	+ve	-ve
2	1/32	+ve	+ve	1/16	+ve	-ve
3	1/256	+ve	+ve	ND	ND	-ve
4	1/512	+ve	+ve	1/128	+ve	-ve
5	1/64	+ve	+ve	1/256	+ve	+ve
6	1/256	+ve	+ve	1/32	+ve	-ve
7	1/512	+ve	+ve	1/256	+ve	-ve



(An ISO 3297: 2007 Certified Organization)

Vol. 3, Issue 6, June 2014

8	1/1024	+ve	+ve	1/512	+ve	-ve
9	1/1024	+ve	+ve	1/512	+ve	-ve
10	1/1024	+ve	+ve	1/1024	+ve	+ve
11	1/1024	+ve	+ve	1/512	+ve	-ve
12	1/64	+ve	+ve	1/512	+ve	-ve
13	1/1024	+ve	+ve	1/512	+ve	-ve
14	1/32	+ve	+ve	1/64	+ve	-ve

IHA: indirect Hemagglutination ELISA: Ezyme linked immunesorbent assay ND: not done +ve: positive, -ve: Negative.

Table 2. Results of serological tests and PCR of a group of non treated Trypanosoma cruzi infected, young adults .

Patie nts	Beginning			Follow up		
	IHA	ELIS	PC	IHA	ELI	PCR
		А	R		SA	
А	1/64	+ve	+ve	ND	ND	+ve
В	1/128	+ve	+ve	1/256	+ve	+ve
С	1/64	+ve	+ve	ND	ND	+ve
D	1/16	+ve	+ve	ND	ND	+ve
Е	1/128	+ve	+ve	ND	ND	+ve

IV. DISCUSSION

The criterion of cure for Chagas disease is still the negativization of conventional serologic and parasitological tests (5), even though there is not much experience using this criterion in chronically infected adults. During this stage of the disease, conventional parasitologic tests display low sensitivity and serological reactions become negative very late, even in cases of successful treatment (1, 10). PCR could overcome both difficulties: it could provide results on the efficacy of treatment with higher sensitivity than conventional parasitological tests, and earlier in time than serological tests. In this work, a significant negativization of PCR was demonstrated in the TG. Although these results are promising for monitoring treatment, the relatively low sensitivity of PCR should be considered. 65.5% of chronically infected patients were positive by PCR. Similar results were observed by other authors (11). A negative result after treatment therefore does not indicate cure, although it could provide a supporting evidence of cure. Only two patients presented a positive PCR after treatment. The possibilities of contamination and lack of treatment compliance were reasonably discarded. Therefore, these results could indicate failure to therapy. This fact has been shown in other studies (2), and could be due to infection with resistant parasite strains (12). An important proportion of adverse effects of benznidazole was observed. However, most of them were mild or could be resolved with simple clinical indications. To evaluate the effectiveness of PCR as method for assessing therapy in chronically infected adults, its sensitivity should be increased and patients should be followed-up for longer periods.



(An ISO 3297: 2007 Certified Organization)

Vol. 3, Issue 6, June 2014

REFERENCES

1. Andrade SG. Tratamiento específico experimental da Doença de Chagas. Rev Patol Trop 29 (supl): 179-189, 2000.

2. Braga MS, Lauria-Pires L, Arganaraz ER, Nascimento RJ, Teixeira AR. Persistent Infections in Chronic Chagas' disease patients treated with anti-Trypanosoma cruzi nitroderivates. Rev Inst Med Trop São Paulo 42: 157-161, 2000.

3. Britto C, Cardoso MAB, Wincker P, Morel CM. A simple protocol for the physical cleavage of Trypanosoma cruzi kinetoplast DNA present in blood samples and it use in polymerase chain

reaction (PCR) - based diagnosis of chronic Chagas disease (Technical note). Mem Inst Oswaldo

Cruz 88: 171-172, 1993.

4. Bronfen E, Rocha FSA, Machado GBN, Perillo MM. Isolamento de amostras do Trypanosoma

cruzi por Xenodianóstico e hemocultura de pacientes na fase crônica da doença de Chagas. Mem

Inst Oswaldo Cruz 84: 237-240, 1989.

5. Cançado R, Brener Z. Terapêutica. In Z Brener, Z Andrade (eds) Trypanosoma cruzi e Doença de Chagas, Guanabara Koogan, Rio de Janeiro, 1979. 6. Florian Sanz F, Gomez Navarro C, Castrillo Garcia N, Pedrote Martinez A, Lage Galle E. Chagasic cardiomyopathy in Spain: a diagnosis to bear in

mind. An Med Interna 22: 538-540, 2005.

7. Kirchoff LV. American trypanosomiasis (Chagas' disease), a tropical disease now in the United States. N Engl J Med 329: 639-644, 1993.

8. Ministério da Saúde. Tratamento Etiológico da Doença de Chagas. 2nd ed., Fundação Nacional de Saúde, Brasilia, 1997.

9. Normas para la atención al paciente infectado con Trypanosoma cruzi (Enfermedad de Chagas). Ministerio de Salud y Medio Ambiente de la República Argentina, 2005.

10. OPS/HCP/HCT/140/99. Tratamiento etiológico de la enfermedad de Chagas: conclusiones de

reunión de especialistas. Rev Patol Trop 28: 247-279, 1999.

11. Portela-Lindoso AA, Shikanai-Yasuda MA. Chronic Chagas' disease: from xenodiagnosis and hemoculture to polymerase chain reaction. Rev Saude Publica 37: 107-115, 2003.

12. Stoppani AOM. Quimioterapia de la enfermedad de Chagas. Medicina (Buenos Aires) 59 (Supl II): 147-65, 1999.

13. Sturm NR, Degrave W, Morel C, Simpson L. Sensitive detection and schizodeme classification of Trypanosoma cruzi cells by amplification of kinetoplast minicircle DNA sequences: use in diagnosis

of Chagas' disease. Mol Biochem Parasitol 33: 205-214, 1989.

14. Wincker P, Britto C, Borges Pereira J, Cardoso MA, Oeleman W, Morel C. Use of a simplified polymerase chain reaction procedure to detect Trypanosoma cruzi in blood samples from chronic chagasic patients in a rural endemic area. Am J Trop Med Hyg 51: 771-777, 1994.

15. World Health Organization. Control of Chagas Disease. Tech Rep Ser 905: 1-109, 2002