

ISSN(Online) : 2319-8753 ISSN (Print) : 2347-6710

International Journal of Innovative Research in Science, Engineering and Technology

(An ISO 3297: 2007 Certified Organization)

Website: <u>www.ijirset.com</u> Vol. 6, Issue 1, January 2017

Effect of Imidacloprid on Haematological Parameters of a *Channa Punctatus* (Bloch.) And Reversal Responses

Pawara, Ravindra H¹, Patel, Nisar G².

P.G. and Research Centre, Department of Zoology, Pratap College, Amalner, Dist-Jalgaon, (M.S.), India¹

Bapumiya Sirajoddin Patel ACS College, Pimpalgaon Kale, Dist- Buldhana, (M.S.), India²

ABSTRACT: The objective of this study was to investigate the reference values for haematological ranges and recovery responses for freshwater teleost *Channa punctatus* (Bloch.). The fish was exposed to sublethal concentration (0.282 ppm) for 24, 48, 72 and 96 hrs to assess the alterations in the level of haematological parameters for acute study. Significant changes in haematological parameters such as RBCs, WBCs and Hb of fish, *Channa punctatus* were observed. These changes were found to be time and dose dependent.

KEYWORDS: Haematological parameters, Imidacloprid, Channa punctatus.

I. INTRODUCTION

Environmental pollution problem in the developing world are clearly linked to unbalanced ecosystem. Environment contamination by pesticides has been documented in both biotic and abiotic components. The random use of different pesticides often causes lot of damage to non target organism. The pollution of environment due to use of pesticides has become an increasing problem over the last century with the development of industry, agriculture and increase in population (Syed, et al., 2012). Several workers have studied haematological alterations due to the pesticide toxicity in fish (Kumar and Nelson, 1997; Tilak, et al., 2003; Shukla, et al., 2005 and Remia, et al., 2008). Modern agricultural practices result in indiscriminate use of various pesticides, which usually enter into the aquatic environment. The use of pesticides in the field has the potential to change the aquatic medium, affecting the tolerance limit of aquatic fauna and flora, as well as creating danger to the ecosystem (Adedeji and Okocha, 2012). Water pollution has emerged as one of the most pressing problems of this century. Water quality management, water pollution control and environmental protection are the main issues to save our future. Imidacloprid (Confidor) is one of the most widely used insecticides in India and other parts of the World. It is sold under many trade names which include Confidor, Admire, Gaucho, Merit, Advantage, Provado, and Winner. The International Union of Pure and Applied Chemistry (IUPAC) name is 1-(6-chloro-3-pyridylmethyl)-N-nitroimidazolidin-2-ylideneamine and chemical formula is C₉H₁₀ClN₅O₂ (CCME, 2007). Neonicotinoid insecticides are grouped into 1) Nitroguanidine: Clothianidin, Dinotefuran, Imidacloprid, Imidaclothiz, Thiamethoxam. 2) Nitromethylene: Nitenpyram, Nithiazine. 3) Pyridylmethylamine: Acetamiprid, Imidacloprid, Nitenpyram, Paichongding, Thiacloprid. 4) Nereistoxin analogue insecticides: Bensultap, Cartap, Polythialan, Thiocyclam, Thiosultap.

The present study has been undertaken to have a basic understanding of the acute toxicity of Imidacloprid insecticide on the experimental fish *Channa punctatus*. Imidacloprid enters the target pest via ingestion or direct contact. Imidacloprid is used for controlling sucking insects, soil insects, termites, and some chewing insects. India is a major supplier of fish in the world. In 2006 the country exported over 6, 00,000 metric tonnes of fish, to some 90 countries, earning over \$1.8 billion. In 2008 India was the sixth largest producer of marine and freshwater capture fisheries, and the second largest aquaculture farmed fish producer in the world (CIFT, 2008). Man is using a number of chemicals and there are a number of chemicals in the environment now. Some of the chemicals are toxic to living system which get incorporated into food chain, disturb the biochemical processes occurring in the living body causing deformities, diseases and even death are called environmental toxicants (Patnaik, *et al.*, 2010). India is an agricultural



(An ISO 3297: 2007 Certified Organization)

Website: www.ijirset.com

Vol. 6, Issue 1, January 2017

country with a large rural population (60-80%), where pesticides are freely available and are used extensively. Haematological tests are important diagnostic tools and recent findings have suggested that they may be equally valuable as indicators of disease or stress due to pollutants and environmental fluctuations in fish. Haematological parameters are related to responses of the organisms to the changing environmental conditions and therefore can be used to screen the healthy state of fish experimented to the exposed toxicant.

II. MATERIAL AND METHODS

Healthy fresh water fish, Channa punctatus (av. length 08.40±1.14 to 10.50 ±1.56 cm and av. weight 12.14±0.45 to 15.54±0.48 gm) of both sexes were purchased from local market of Amalner, district, Jalgaon and used as the test animal, carefully brought to the laboratory avoiding any injury to the fish during transport. Identification of fish was done with the help of standard keys and available literature. The fish were disinfected with 0.01% solution of KmNO₄ for two minutes and then transferred to a 45 litre ($60 \times 30 \times 45$ cm) capacity glass aquarium to acclimatized for 15 days in laboratory conditions before use. The tank water was aerated continuously and food was provided daily in the morning in the form of chopped goat liver, only white part of boiled egg, meat, chicken as well as pieces of earthworm. After acclimatization, fish were kept in different concentrations of Imidacloprid in to 5 litre capacity plastic tubs. Finney (1971) Probit Analysis LC_{50} determination method was used to find out the LC_{50} concentration. The formula used to assess the mortality of the fish recommended by the WHO (2004) and FAO (2002). The commercial grade Imidacloprid (1-[6-chloro-3-pyridylmethyl]-N-nitroimidazolidin-2-ylideaneamine) was chosen for the experiment and was obtained from the local market, Amalner. The toxicity of the Imidacloprid on the fresh water fish, Channa punctatus was assessed and determined LC₁₀, LC₅₀ and LC₉₀ for 24, 48, 72 and 96 hrs by probit analysis as per Organisation for Economic Co-operation and Development (OECD) guidelines for testing of chemicals (OECD 203 "Fish, acute toxicity test", 1992 and 2010). After completion of experimental and control groups, five fish were taken without anesthesia after an interval of 24, 48, 72 and 96 hrs for haematological study and five fish were remained for recovery study. Blood was collected from five fish from each group by heart puncturing as well as posterior caudal vein using Ethylene-diaminetetraacetate (EDTA) as anticoagulant. Red blood corpuscles (RBCs), White blood corpuscles (WBCs) counts were done with an improved Neubauer haemocytometer (Shah and Altindag, 2004) under the Microscope (Olympus Microscope 20i BIMF, Olympus (India) Pvt, Ltd, Noida) and Hemoglobin (Hb) concentration was measured by Sahli's method (Sahli, 1962). The results obtained were subjected to analysis for mean and standard deviation. The mean values of treatment were subjected to statistical analysis using two-way analysis of variance (ANOVA) to test the (0.05) level of significance between the various sublethal concentrations of Imidacloprid.

III. RESULT AND DISCUSSION

In present study White Blood corpuscles increased, red blood corpuscles counts exhibited a decrease in their numbers and hemoglobin values also decreased significantly were reported after exposure to the sublethal dose of Imidacloprid. Malathi, et al. (2012) studied comparatively ranges values of haematological parameters in C. punctatus and C. striatus. They reported as RBC 3.29×10^3 to 3.42×10^6 /cumm; WBC 4.3 to 4.6×10^3 /cumm and haemoglobin 9.37 to 10.84 g/dl. In present study normal values of haematological parameters were reported as RBC 3.28×10^{6} /µl; WBC 14.44×10^{3} /µl and haemoglobin 13.1 g/dl. RBCs decreased to 2.67×10^{6} /µl (18.59%), 2.33×10^{6} /µl (28.96%), $2.19 \times 10^{6}/\mu$ (33.23%) as well as $2.47 \times 10^{6}/\mu$ (24.69%) respectively after 24 hrs, 48 hrs, 72 hrs and 96 hrs respectively and 3.066×10^{6} /µl (6.52%) recovered after 10 days (Fig. 1.1). Whereas in experimental group WBCs mean level increased to 42.32±2.174 (193.07%), 31.38±3.356 (117.31%), 62.16±4.926 (330.47%) as well as 55.14±1.443 (281.85%) for 24 hrs, 48 hrs, 72 hrs, and 96 hrs for sub lethal exposure of Imidacloprid respectively and after 10 days recovery 16.2±3.043 (12.18%) (Fig. 1.3) and The fish were exposed to sublethal concentration of Imidacloprid showed the haemoglobin mean values for 24 hrs, 48 hrs, 72 hrs as well as 96 hrs 11.42±0.356 g/dl (12.82%), 10.96±0.55 g/dl (16.33%), 10.4±0.223 g/dl (20.61%) and 11.3±0.839 g/dl (13.74%) respectively and after 10 days recovery mean value 12.9±0.353 g/dl (1.526%) (Fig. 1.5). The percentage increase/decrease of RBCs, WBCs and Hb in various time exposures for 24, 48, 72 and 96 hrs and recovery values were showed in figure 1.2, 1.4 and 1.6 respectively. Variations and percent increase/decrease in Red blood corpuscles (RBCs), WBCs and Haemoglobin count



(An ISO 3297: 2007 Certified Organization)

Website: www.ijirset.com

Vol. 6, Issue 1, January 2017

of control and treated fish with Imidacloprid are depicted in table.1.1.1, 1.2.1 and 1.3.1 respectively. Comparison of data of control with that of the treated groups very clearly indicates. Reduction of RBC in treated groups at sublethal concentrations of Imidacloprid (0.282ppm). RBC count value decreased significantly (p < 0.05) as compared to control group. After 10 days recovery period RBCs count was observed normally as compared to control fish.







(An ISO 3297: 2007 Certified Organization)

Website: www.ijirset.com

Vol. 6, Issue 1, January 2017

 Table 1.1.1. The variation in RBC, WBC and Hb count of *Channa punctatus* exposed to sublethal concentration of Imidacloprid.

Exposure period (hrs)	RBC (10 ⁶ /µl)	Percent decrease
Control	3.28±0.461	-
24	2.67±0.157*	18.59*
48	2.33±0.129*	28.96*
72	2.19±0.106*	33.23*
96	2.47±0.152*	24.69*
Recovery	3.066±0.358	6.52

Data are represented as mean±SD.

Data were analyzed through two-way ANOVA.

*Significant (P< 0.05), when treated groups were compared with control.





(An ISO 3297: 2007 Certified Organization)

Website: www.ijirset.com

Vol. 6, Issue 1, January 2017



Table 1.2.1: The variation in WBC of *Channa punctatus* exposed to sublethal concentration of Imidacloprid.

WBC	Percent
$(10^{3}/\mu l)$	increase
14.44±0.661	-
42.32±2.174*	193.07*
31.38±3.356*	117.31*
62.16±4.926*	330.47*
55.14±1.443*	281.85*
16.2±3.043	12.18
	(10³/µl) 14.44±0.661 42.32±2.174* 31.38±3.356* 62.16±4.926* 55.14±1.443*

Data are represented as mean±SD.

Data were analyzed through two-way ANOVA.

*Significant (P < 0.05), when treated groups were compared with control.





(An ISO 3297: 2007 Certified Organization)

Website: www.ijirset.com

Vol. 6, Issue 1, January 2017



Table 1.3.1: The variation in Haemoglobin of Channa punctatus exposed to sublethal concentration of Imidacloprid.

Exposure period	Hb	Percent
(hrs)	(g/dl)	decrease
Control	13.1±0.387	-
24	11.42±0.356*	12.82*
48	10.96±0.55*	16.33*
72	10.4±0.223*	20.61*
96	11.3±0.839*	13.74*
Recovery	12.9±0.353	1.526

Data are represented as mean±SD.

Data were analyzed through two-way ANOVA.

*Significant (P<0.05), when treated groups were compared with control.

An increase in the WBCs count was mostly observed during the exposure period of stress reaction when fish tried to restore the disturbed homeostasis. High white blood cell counts indicate damage due to infection of body tissues, severe physical stress, and as well leukemia. The value mentioned above showed a significant decrease RBCs, Hb and increase WBCs when compared with control (p < 0.05). Similar level as a control fish were observed in the C. punctatus after recovery period. Padma, et al. (2012) reported different concentrations (0.002 ppm, 0.004 ppm, 0.006 ppm, 0.008 ppm and 0.010 ppm) of Imidacloprid insecticide on serum biochemical parameters of fresh water teleost Channa punctatus. Saxena and Seth (2002) noticed the toxic effects of Cypermethrin on certain hematological aspects of fresh water fish Channa punctatus and observed a significant change in the hematology of the fish. Anees (1978) observed a decrease in erythrocyte count and haemoglobin content in freshwater fish Channa punctatus after acute exposure to Diazinon. Singh, et al. (2008) showed effect of Copper on haematological profile of freshwater fish, Channa punctatus a significant decrease in the haemoglobin content from 10.73 to 6.60%, red blood cells from 2.86 to 1.84×10^{6} /mm³ and packed cell volume from 31.00 to 23.33%. Whereas the white blood cells increased from 60.00 to 92.48 x10³/mm³ at the end of 45th day as compared to control. Reddy, *et al.* (2015) obtained the static LC₅₀ values were lethal (14,260 mg/l) and 13,710 mg/l and 11,900 mg/l for 24 and 96 hrs respectively in fresh water fish Labeo rohita (Hamilton) exposed to Confidor (Imidacloprid ingradient). Therefore, present results are in agreement with the observations of the previous workers.



(An ISO 3297: 2007 Certified Organization)

Website: <u>www.ijirset.com</u>

Vol. 6, Issue 1, January 2017

IV. CONCLUSION

Decreased Hb%, RBC count and increased WBC counts indicate damage due to infection of body tissues, severe physical stress, and as well leukemia. In most cases, abnormal red cell morphology is noted. Haematological parameters related to Oxygen transport (RBC and Hb), defense mechanisms (WBC) all exhibited marked alterations between control and experimental groups in response to insecticide Imidacloprid in presently studied fish, *C. punctatus*. Present studies thus confirm that haematological parameters are very sensitive indicators in fish under toxicity of chemicals and ultimately become the causative for affecting the general health status of the fish. Water quality can thus be regarded as one of the major factors responsible for individual variations in fish haematology.

ACKNOWLEDGEMENTS

Authors are thankful to The Chairman, K.E. Society; Prof. Dr. Chavhan M. B., Principal, Pratap College, Amalner for continuous encouragement as well as providing excellent library as well as laboratory facilities during the tenure of this work. Thanks are also due to Assitt. Prof. Nitin Patil for helping in strengthening the language. Mr. R. H. Pawara is grateful to UGC, New Delhi for providing Rajiv Gandhi National Fellowship as SRF.

REFERENCES

- 1. Adedeji, O.B. and Okocha, R.O. (2012). Overview of pesticide toxicity in fish. Advances in Environmental Biology, 6(8): 2344-2351.
- Anees, M.A. (1978). Haematological abnormalities in a freshwater teleost, *Channa punctatus* (Bloch.), exposed to sublethal and chronic levels of three organophosphorus insecticides. Int. J. Ecol. Environ. Sci., 4: 53-60.
- 3. Canadian Council of Ministers of the Environment, (CCME). (2007). Canadian water quality guidelines for the protection of aquatic life: Imidacloprid. Scientific Supporting Document. Winnipeg, Manitoba., 1-8.
- 4. Central Institute of Fisheries Technology, (CIFT). (2014). Annual Report 2013-14, CIFT Junction, Matsyapuri P.O., Cochin 682 029.
- 5. Finney, D.J. (1971). Probit analysis. 3rd edition. Cambridge University Press, London and New York.
- 6. Kumar, G.S. and Nelson, D.J. (1997). Effect of Ekalux on haematology of the fresh water fish *Etroplus maculates*. J. Ecotoxicol. Environ. Monit., 7: 113-116.
- 7. Malathi K., Kannathasan, A. and Rajendran, K. (2012). Comparative haematological studies on fresh water fishes *Channa punctatus* and *Channa striatus* (Bloch.). IJPCBS., 2(4): 644-648.
- Organisation for Economic Cooperation and Development, (OECD) (1992). Test guideline 203, OECD guideline for testing of chemicals, fish, acute toxicity test.
- 9. Organisation for Economic Cooperation and Development, (OECD) (2010): Short guidance on the threshold approach for acute fish toxicity. Series on testing and assessment No. 126, ENV/JM/TG(2010)/7, OECD, Paris.
- 10. Padma, Priya B., Rachel, Vijaya, Maruti, Y.A. (2012). Acute toxicity effect of Imidacloprid insecticides on serum biochemical parameters of fresh water teleost *Channa punctatus* (Bloch.). International Journal of Integrative sciences, Innovation and Technology, 1(2): 18-22.
- 11. Reddy, A., Veeraiah, K. Tata, Rao. S. and Vivek, Ch. (2015). Studies on some biochemical changes in the tissues of the fresh water fish *Labeo* rohita (Hamilton) exposed to Confidor. Journal of International Academic Research for Multidisciplinary, 3(1): 323-329.
- 12. Remia, K.M., Logankumar, S. and Rajmohan, D. (2008). Effect of an insecticide (Monocrotophos) on some biochemical constituents of the fish *Tilapia mossambica*. Poll. Res., 27(3): 523-526.
- 13. Sahli (1962). Determination of haemoglobin by acid haematin method. In: Practical haematology (Ed: Dacie J.V. and Lewis S.M.) 5th Edn. Churuchill, London.
- 14. Saxena, K.K. and Seth, N. (2002). Toxic effects of Cypermethrin on certain hematological aspects of fresh water fish *Channa punctatus*. Bull. Environ. Contam. Toxicol., 69: 364-369.
- 15. Shah, S.L. And Altindag, A. (2004). Haematological parameters of tench (*Tinca tinca* L.) after acute and chronic exposure to lethal and sublethal Mercury treatments. Bull. Environ. Contam. Toxicol., 73: 911-918.
- 16. Shukla, Smita, Dass, K. Seyata and Saksena, D.N. (2005). Biochemical alteration in the liver and kidney of a freshwater catfish *Clarius batrachus* (Linn) exposure of organophosphorus pesticides Nuvan (DDVP). J. Aqua. Biol., 20(1): 135-140.
- 17. Singh, D., Nath, K., Trivedi, S.P. and Sharma, Y.K. (2008). Impact of Copper on haematological profile of freshwater fish, *Channa punctatus*. Journal of Environmental Biology, 29(2): 253-257.
- 18. Syed, A., Fathima, K.M., Annalakshmi, T. and Xavier, I. B. (2012). Immunostimulant effect of vitamin-A in *Channa Punctatus* challenged with Aeromonas Hydrophila: haematological evaluation. Journal of Applied Pharmaceutical Science, 2(11): 123-126.
- 19. Tilak, K., Sathyavardhan, S.K. and Thathaji, P.B. (2003). Biochemical changes induced by Fenvalerate in the fresh water fish *Channa punctatus*. J. Ecotoxciol. Environ. Monit., 13(4): 261-270.